Possible acquired resistance of dogs successively infested by *Amblyomma cajennense* (Fabricius, 1787) (Acari: Ixodidae) nymphs

Possibilidade de ocorrência de resistência adquirida em cães sucessivamente infestados por ninhas de *Amblyomma cajennense* (Fabricius, 1787) (Acari: Ixodidae)

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Abstract

The present study aimed to evaluate the occurrence of immune resistance in dogs successively infested with *Amblyomma cajennense* nymphs. Five animals were submitted to four consecutive infestations with *A. cajennense* nymphs, at fourteen-day intervals. For each infestation, 50 nymphs were used per animal and data on the parasitic and non-parasitic periods were recorded. The average recovering rate of engorged nymphs in the successive infestations were 52.0, 29.2, 9.6 and 12.8%, respectively, with a significant reduction (p < 0.05) of this parameter from the second infestation onwards. The modal drop-off day of engorged nymphs was Day 4 of parasitism in all infestations. The average mortality rates of nymphs seen on the first, second, third and fourth infestations were 3.6, 3.2, 2.0 and 2.8%, respectively, with no significant differences among them (p < 0.05). In addition, no significant differences were seen among the ecdysis rates for specimens recovered from successive parasitic challenges. The study results suggest that the acquired resistance of infested dogs had a negative effect on recovery rate of *A. cajennense* nymphs; however, it did not affect the other biological parameters evaluated.

Keywords: Immune resistance, *Amblyomma cajennense*, Ixodidae, dogs.

Resumo

O presente trabalho teve por objetivo avaliar a ocorrência de resistência imune em cães, frente a infestações sucessivas por ninhas de *Amblyomma cajennense*. Para tanto, cinco animais foram submetidos a quatro infestações consecutivas por ninhas de *A. cajennense* em intervalos de quatorze dias. Foram aplicadas 50 ninhas em cada animal por infestação e os dados referentes aos períodos parasitários e não parasitários, foram registrados. As taxas médias de recuperação de ninhas ingurgitadas, verificadas nas sucessivas infestações foram de 52,0, 29,2, 9,6 e 12,8%, sendo observada uma redução significativa (p < 0,05) nesse parâmetro a partir da segunda infestação. O dia modal de queda das ninhas ingurgitadas em todas as infestações foi o 4º dia de parasitismo. As taxas médias de mortalidade de ninhas observadas no primeiro, segundo, terceiro e quarto desafio parasitário foram, respectivamente, 3,6, 3,2, 2,0 e 2,8%, não havendo diferença significativa entre elas (p < 0,05). Não observou-se diferença significativa (p < 0,05) entre as taxas de ecdise reportadas para os exemplares recuperados nos sucessivos desafios parasitários. Esses resultados sugerem que a resistência adquirida nos cães parasitados afetou negativamente a taxa de recuperação das ninhas de *A. cajennense* inoculadas nesses animais, contudo não apresentou nenhum efeito sobre os demais parâmetros biológicos avaliados.

Introduction

In tropical regions, a vast variety of ticks infest dogs living either in urban areas, where *Rhipicephalus sanguineus* is the predominant species (SZABÓ et al., 2001; OYAFUSO et al., 2002), or in peri-urban and rural areas, where dogs are frequently infested with *Amblyomma* species, including *Amblyomma cajennense* (LABRUNA et al., 2000). This tick is popularly known as “star tick” or “rodoleiro” tick. Direct damages caused by this species in infested animals include cutaneous injuries, allergic reactions, blood loss and the potential to transmit infectious agents (O’DWYER et al., 2001). Furthermore, it should be emphasized its public health role, as *A. cajennense* is the main ixodide population infesting human beings. This tick species is a potential vector of *Rickettsia rickettsii*, the etiological agent of Rocky Mountain spotted fever in neotropical areas (GUEDES et al., 2005).

Despite all direct and indirect damages caused to domestic animals and man, the development of resistance in these hosts due to the parasitic challenge is likely to occur, through effector mechanisms of innate and acquired immune responses (CRAIG et al., 1996, BROSSARD; WIKEL, 2004). Such resistance develops after the first tick exposure and is, expressed in subsequent infestations by a decrease in number and body weight of engorged females, a decrease in number and viability of eggs, an increase in the engorgement period and mortality of ixodides during the feeding period on the host (BALASHOV, 1972; BROSSARD; WIKEL, 2004). These phenomena may limit tick ability to transmit infectious agents to their hosts, as they interfere directly in the parasitic process.

In a pilot study carried out in our laboratory in which dogs were repeatedly infested with *A. cajennense* nymphs, a certain level of resistance was seen after the second infestation onwards. These data contrasts with that reported in a similar study by Mukai et al. (2002) who did not find, this same immune resistance profile in dogs infested with *A. cajennense* nymphs.

Considering the close contact between man and dogs and the potential of dogs to disseminate ticks and consequently transmit diseases with zoonotic potential, it is important to describe the aspects involved in the interaction between dogs and *A. cajennense* nymphs. Thus, the present study aimed to evaluate the occurrence of immune resistance in dogs submitted to successive infestations by *A. cajennense* nymphs.

Material and Methods

The study was carried out at the Veterinary School, Federal University of the state of Minas Gerais, Brazil, from April to October 2005. The experiment was developed at the laboratory of endo and ectoparasitosis of the Department of Preventive Veterinary Medicine (DMVP) and DMVP animal nursery.

Five-months-old mongrel dogs (two females and three males), from two offsprings born in Belo Horizonte, Minas Gerais, were used for this experiment. Before initiating the experiment all dogs were vaccinated against hepatitis, distemper, parvovirus and leptospirosis, and were treated with antihelminthic drugs. The study animals were kept isolated throughout the experimental period at DMVP animal nursery. They were kept in especially designed individual wire cages of 50 × 60 × 120 cm. The cages were kept 40 cm above the floor in order to allow animal hygiene and keep the environment dry and comfortable. Water and commercial feed were offered ad libitum. The room was cleaned twice a day and included washing food and water containers and removal of feces and urine. In addition, the air-exhaust ventilation units were kept on for good air flow.

The ticks used in this experiment were obtained from two manual collections of engorged larvae from the body of an artificially infested horse, as described by Oliveira et al. (2003). These collections were carried out within a two-month interval, so that nymphs of approximately one month old were used to infest the study dogs. Engorged specimens were placed into plastic 3 mL plastic syringes that were previously prepared for the inoculation process. Syringes containing 50 engorged larvae were sealed with hydrophilic cotton and placed in an air-conditioned incubator (27 °C and 80% relative humidity), and were monitored daily until completion of ecdysis. Only *A. cajennense* nymphs were used in the experiment.

Each dog was submitted to four consecutive infestations of 50 *A. cajennense* nymphs each, at fourteen-day intervals. On the day of infestation each animal was transferred to a 30 × 40 × 50 cm cage that was placed on a metallic tray, lined by double-coated adhesive tape to prevent dispersion of ticks. For infestation procedure nymphs was released on a piece of nylon carpet that was left inside the cage for 2 hours, following the modified technique as described by Young et al. (2003). After the required time for tick fixation, each dog was put back into its original cage. The trays and carpets were removed from the infestation cages and non-attached ticks were collected and counted.

After each infestation, the animals were monitored daily until complete drop-off of all ticks. Monitoring was carried out twice a day through visual inspection of attached parasites and manual collection of the detached ones. During this period, metallic trays lined by double-coated adhesive tape were placed under each dog’s cage to recover naturally detached ticks. These trays were also inspected twice a day before the cages were cleaned up. The recovered nymphs were placed into Petri dishes, which were incubated as described before (27 °C and 80% relative humidity), during the entire ecdysis period. For each infestation the following parameters were evaluated: mortality rate¹, recovery rate of engorged nymphs², modal day of nymph drop-off, and ecdysis rate of recovered nymphs³. Data analysis included ANOVA (analysis of variance) and means were compared through the Student’s t-test at 95% confidence interval (p < 0.05).

Results and Discussion

The biological parameters for recovered nymphs during the successive infestations are shown in Table 1.

The average recovery rates of engorged nymphs after successive infestations were 52.0, 29.2, 9.6 and 12.8%. A significant decrease (p < 0.05) was seen in the mean number of ticks that were able to feed on dogs after the first infestation. This reduction may be

¹Mortality rate = (no. of recovered dead nymphs + no. of inoculated nymphs) × 100
²Recovery rate = (no. of recovered engorged nymphs + no. of inoculated nymphs) × 100
³Ecdysis rate = (no. of nymphs showing ecdysis + no. of recovered engorged nymphs) × 100
explained by the dogs’ immune response on tick fixation sites, which can directly affect the feeding process of ticks or increase the animal’s self-cleaning behavior, as an indirect consequence of that response (CRAIG et al., 1996). These results differ from those reported by Mukai et al. (2002) who did not find a significant difference on recovering rates of engorged A. cajennense nymphs after three successive infestations on dogs. It is possible that these authors’ infestation method, which consisted of placing ticks in feeding chambers that were attached to the animal body, resulted in higher efficiency of inoculated nymphs. In contrast, in the present study, we tried to apply an infestation approach that was as closer as possible to the natural parasitic conditions, allowing natural grooming behavior, as expected in naturally infested animals.

Mean mortality rates of nymphs seen in the first, second, third and fourth infestations were 3.6, 3.2, 2 and 2.8%, respectively, with no significant differences among them (p < 0.05). Similarly, no significant differences were found (p < 0.05) among ecdysis rates for specimens recovered from successive parasitic challenges (79.3, 81.2, 90.0 and 87.6%). According to Balashov (1972), the minimum required weight to enable semi-engorged ixodides to undergo ecdysis corresponds to 20% of their normal weight after engorgement. Thus, probably almost all nymphs recovered in the present experiment were able to feed and reach the minimum required weight to undergo ecdysis to the next stage, even when conditions were not suitable for parasitism. This would explain the fact that ecdysis and mortality rates of recovered nymphs were not influenced by the development of immune resistance during successive infestations.

The modal day for drop-off of engorged nymphs for all infestations was Day 4 of parasitism.

Although ecdysis and mortality rates and the modal day for drop-off of A. cajennense nymphs were not influenced by the successive exposures of hosts to ticks, immune resistance of dogs was apparently the phenomenon causing significant decrease of recovery rates of engorged nymphs either through the development of a local inflammatory response or increasing grooming behavior of infested animals. However, further experiments using naïve dogs in each infestation are needed to confirm the occurrence of resistance; this would standardize and ensure the quality of nymph inocula.

Table 1. Biological parameters (mean ± standard deviation) of A. cajennense nymphs recovered after four successive infestations in dogs (Belo Horizonte, Brazil, 2005).

<table>
<thead>
<tr>
<th>Parameter (%)</th>
<th>First</th>
<th>Second</th>
<th>Third</th>
<th>Fourth</th>
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<tr>
<td>Recovery rate</td>
<td>52.0 ± 6.8&lt;sup&gt;a&lt;/sup&gt;</td>
<td>29.2 ± 14.3&lt;sup&gt;b&lt;/sup&gt;</td>
<td>9.6 ± 5.9&lt;sup&gt;c&lt;/sup&gt;</td>
<td>12.8 ± 6.4&lt;sup&gt;c&lt;/sup&gt;</td>
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<tr>
<td>Ecdysis rate</td>
<td>79.3 ± 6.4&lt;sup&gt;a&lt;/sup&gt;</td>
<td>81.2 ± 12.7&lt;sup&gt;b&lt;/sup&gt;</td>
<td>90.0 ± 22.4&lt;sup&gt;c&lt;/sup&gt;</td>
<td>87.6 ± 14.0&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>Mortality rate</td>
<td>3.6 ± 2.2&lt;sup&gt;a&lt;/sup&gt;</td>
<td>3.2 ± 4.1&lt;sup&gt;c&lt;/sup&gt;</td>
<td>2.0 ± 1.4&lt;sup&gt;c&lt;/sup&gt;</td>
<td>2.8 ± 2.4&lt;sup&gt;c&lt;/sup&gt;</td>
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<sup>a,b,c</sup> Means with no significant difference (p < 0.05).

References


